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Synthesis of a novel glycosphingolipid from the millipede, Parafontaria laminata armigera, and the assembly of its carbohydrate moiety into multivalent structures

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Abstract—A novel glycosphingolipid, β-D-Man*p*- $(1\rightarrow 4)$ -[α-L-Fuc*p*- $(1\rightarrow 3)$]-β-D-Glc*p*- $(1\rightarrow 1)$ -Cer, found in the millipede, *Parafontaria laminata armigera*, and multivalent derivatives of its carbohydrate moiety were synthesized. As the key step, the target glycolipid (1) was obtained through an inversion reaction at the 2-position of a β-glucopyranoside residue yielding a β-mannopyranoside. In addition, the synthesis of fluorescently labeled trimer and tetramer glycoconjugates (2, 3) was achieved by iterative amide bond formation using a monomer unit (24).

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1. Introduction

In our continuing studies to elucidate the biological function of glycoconjugates, we have synthesized novel glycosphingolipids found in various invertebrates $^{1-6}$ that do not have gangliosides. A number of glycosphingolipids from a variety of invertebrate origins and related analogues have been synthesized and their influence on proliferation has been examined using mouse melanoma B16 cells. In these studies, the glycolipid, β -D-Manp- $(1\rightarrow 4)$ - $[\alpha$ -Fucp- $(1\rightarrow 3)]$ - β -D-Glcp- $(1\rightarrow 1)$ -Cer, which has been isolated from the millipede *Parafontaria laminata armigera*, has been observed to inhibit significantly cell proliferation (details will be reported elsewhere).

In a previous paper,² we reported the synthesis of this glycosphingolipid. The key reaction in that synthesis was a two-step glycosylation. The first step was the formation of an orthoester from two sugar moieties and the second step was the reductive cleavage of one of the orthoester C–O bonds leading to β -selective mannosyl-

ation. This earlier paper was the first report on the total synthesis of β -D-Manp- $(1\rightarrow 4)$ - $[\alpha$ -Fucp- $(1\rightarrow 3)]$ - β -D-Glcp- $(1\rightarrow 1)$ -Cer by this method. Among β -mannopyranoside syntheses, one approach is the so called intermolecular nucleophiles approach, in which an inversion reaction at the 2-position of β -glucopuranoside yields a β -mannopyranoside. The strategy, which provides good yields, relies upon creating a highly reactive leaving group at the position that is to be inverted and then treating it with a strong nucleophile. The nucleophile causes $S_N 2$ displacement and epimerizes the position on the carbohydrate ring. We report here the application of this approach for the synthesis of glycosphingolipid 1.

It is known that oligosaccharide chains generally interact with their protein receptors in a multivalent fashion to overcome the inherently low affinity of monovalent carbohydrate—protein interactions. Therefore, the construction of a clustered glycoconjugates is an important subject in glycoscience. ^{11,12} For this reason, we have used a new method to synthesize new peptidic glycoclusters and glycodendrons (2, 3), which consist of a β -alanine derivative linked to the sugar moiety of 1 (Fig. 1). ^{13,14}

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Figure 1. Structure of target glycoconjugates.

2. Results and discussion

2.1. Syntheses of monosaccharide derivatives

Syntheses of the glucopyranose building blocks **6a** and **8** were carried out as depicted in Scheme 1. Compound **6a** was prepared from known 2-(trimethylsilyl)ethyl 4,6-*O*-benzylidene-β-D-glucopyranoside (**4**)¹⁵ via successive mono-alkylation of the diol, acylation and reductive ring opening of the 4,6-acetal. Unfortunately, alkylation of **4** via a stanylene intermediate did not proceed in a regiospecific manner. Although the formation of dialkylated product was avoided, the two regioisomeric monoalkylated products could not be separated, nor could their benzoylated derivatives **5a** and **5b** be separated. The structures of **5a** and **5b** were confirmed after reductive ring opening of the benzylidene acetal in **5**, which yielded a 1.5:1 ratio of **6a** and **6b** in a combined yield of 72%.

Glucopyranosyl donor **8** was obtained from phenyl 4,6-O-benzylidene-1-thio- β -D-glucopyranoside (7), ¹⁶ via a two-step procedure. First, reaction of the stannylidene derivative of **7** with benzyl bromide was found to be highly regiospecific in this case, which may be associated with the electron donating nature of the sulfur aglycon. Second, subsequent chloroacetylation of **7** afforded **8** (81% over two steps).

2.2. Synthesis of the target glycosphingolipid

Glycosylation of acceptor **6a** with **8** in the presence of *N*-iodosuccinimide (NIS), trifluoromethanesulfonic acid (TfOH), ¹⁷ and 4 Å molecular sieves in dichloromethane gave the desired disaccharide (**9**) in 57% yield after purification. The stereochemistry of the newly formed glycosidic linkage could be determined by ¹H NMR spectroscopy (H-1', 4.61 ppm, J = 7.9 Hz). Selective

Scheme 1. Reagents: (a) (i) *n*-Bu₂SnO, benzene; (ii) *n*-Bu₄NBr, *p*-MBnCl, toluene; (b) BzCl, pyridine, 71% two steps; (c) NaBH₃CN, HCl/Et₂O, THF, 72%; (d) (i) *n*-Bu₂SnO, benzene; (ii) *n*-Bu₄NBr, BnBr, toluene; (e) ClAcCl, CH₂Cl₂/pyridine, 81% two steps.

removal of the chloroacetyl group in 9 with NaHCO₃ gave 10, a disaccharide containing a β-D-glucopyranosyl residue with the C2' hydroxyl group unsubstituted. Compound 10 was converted into the triflate by treatment with trifluoromethanesulfonic anhydride in the presence of pyridine at 0 °C. The triflate was then subjected to a nucleophilic substitution reaction with tetrabutylammonium benzoate, which inverted the C2' stereocenter converting the β-p-glucopyranosyl moiety to a β-D-mannopyranosyl residue (11) in good overall yield from 10 (80%). The anomeric hydrogen atom of the mannopyranosyl unit appeared in the NMR spectrum as a signal at δ 4.79 (s, H-1'). The β -D configuration of the glycosidic bond was indicated by the $^{1}J_{\text{C1,H1}}$ value of 157.3 Hz (C-1', 99.4 ppm) in the ^{13}C NMR spectrum. 18 Removal of the 3'-*O-p*MBn group from 11 by DDO gave the disaccharide acceptor 12. Glycosylation of 12 with known fucopyranosyl donor 13¹⁹ in the presence of NIS, TfOH and 4 Å molecular sieves in dichloromethane at -60 °C gave the desired trisaccharide 14 (99%). The stereochemistry of the introduced fucopyranosyl residue was evident by ¹H NMR spectroscopy (H-1", 5.37 ppm, J = 3.7 Hz). Removal of the benzylidene and benzyl group from 14 by catalytic hydrogenolysis over 10% Pd/C in THF/CH₃OH/AcOH (3:3:1) and subsequent acetylation gave 15. Selective removal of the 2-(trimethylsilyl)ethyl (SE) group with trifluoroacetic acid in dichloromethane, and treatment²⁰ with trichloroacetonitrile in the presence of DBU gave the corresponding α -trichloroacetimidate **16**. Glycosylation of (2S,3R,4E)-3-O-benzoyl-2-hexadecanamido-4-octadecene-1,3-diol **17**⁶ with **16** was carried out in the presence of trimethylsilyl trifluoromethanesulfonate (TMSOTf) and 4 Å molecular sieves to afford the desired β -glycoside **18** in 35% yield. Finally, removal of acyl groups in **18** under Zemplen conditions and column chromatography on Sephadex LH-20 furnished a target glycolipid **1** (Scheme 2). The structure and purity of **1** was demonstrated by 1 H NMR spectroscopy and high-resolution fast-atom-bombardment mass spectrometry.

2.3. Syntheses of multivalent glycoconjugates

We planned to use glycosyl donor 16 in the syntheses of the multivalent glycoconjugates. Elongation of the glycocluster was done using three iterative reactions: amide bond formation, deprotection of the *tert*-butoxycarbonyl (Boc) group, and deprotection of the trichloroethyl ester (Tce) group. The required building block 24 was prepared as follows. Coupling of trisaccharide imidate 16 with the spacer 19, which was obtained from hexanolamine, in the presence of TMSOTf gave compound 20 in quantitative yield. Selective removal of the Troc group from 20 by zinc metal and acetic acid gave the primary amine 21 also in quantitative yield. Compound 21 was condensed in the presence of diethyl

Scheme 2. Reagents: (a) NIS, TfOH, 4 Å molecular sieves, CH₂Cl₂ 9: 57%, 14: 99%; (b) NaHCO₃, THF, 76%; (c) (i) Tf₂O, CH₂Cl₂/pyridine; (ii) *n*-Bu₄NBz, DMF, 80%; (d) DDQ, CHCl₃/H₂O (19:1) 78%; (ii) *n*-Bu₄NBr, BnBr, toluene; (e) (i) H₂, Pd/C, CH₃OH/AcOH; (ii) Ac₂O/pyridine, 60%; (f) (i) TFA, CH₂Cl₂; (ii) CCl₃CN, DBU, CH₂Cl₂, 98%; (g) TMSOTf, 4 Å molecular sieves, CH₂Cl₂ 35%; (h) NaOCH₃, 1,4-dioxane/CH₃OH, 94%.

phosphorocyanidate (DEPC) in dry DMF with β-alanine derivative 22, which was prepared according to a previous paper, 13 to give 23 in 66% yield. Subsequent removal of the Tce group with zinc metal and acetic acid afforded an 87% yield of 24, which has a free carboxylic acid moiety. Coupling of 24 with 21 gave the dimer 25 in 71% yield. The Boc group of 25 was removed with 50% TFA affording 26 (86% yield), which was subsequently subjected to the next cycle for the elongation to give the desired trimer glycocluster derivative 27 (79%). Dansyl glycine was introduced into the trimer 28 in the presence of DEPC to give compound 29 in 85% yield. Finally, complete removal of the acyl groups provided the fluorescently labeled glycocluster 2 in 91% yield (Scheme 3). Furthermore, coupling of 28 with 24 in a subsequent cycle gave the desired tetramer derivative 30 in 83% yield, and following the cleavage of the Boc group (56% yield), the resulting amine was coupled with dansyl glycine as described for 29 to give compound 32 in 94% yield. Complete removal of the acyl groups and esters afforded 3 in 91% yield (Scheme 4). The structure and purity of 2 and 3 were supported by ¹H and ¹³C NMR as well as mass spectrometry data.

In conclusion, the synthesis of oligosaccharides containing the β -D-Manp-(1 \rightarrow 4)-Glcp structural motifs has been carried out. In addition, an efficient synthetic strategy was developed to obtain new multivalent glyco-

conjugates. This method should be of use in the preparation of a number of other such species. The results of the biological testing of these compounds will be reported in detail elsewhere.

3. Experimental

3.1. General methods

Optical rotations were measured with a Jasco digital polarimeter. ¹H NMR and ¹³C NMR spectra were recorded with a JMN A500 FT NMR spectrometer with (CH₃)₄Si as the internal standard for solutions in CDCl₃. MALDI-TOF mass spectra (MALDIMS) were recorded on a Perseptive Voyager RP mass spectrometer. High-resolution mass spectra were recorded on a JEOL JMS-700 under FAB conditions (FABMS). TLC was performed on Silica Gel 60 F₂₅₄ (E. Merck) with detection by quenching of UV fluorescence and by charring with 10% H₂SO₄. Column chromatography was carried out on Silica Gel 60 (E. Merck). 2-(Trimethylsilyl)ethyl 4,6-O-benzylidene-β-D-glucopyranoside (**4**), ¹⁵ phenyl 4,6-*O*-benzylidene-1-thio-β-D-glucopyranoside (**3**), ¹⁶ phenyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside (13)¹⁹ and (2S,3R,4E)-3-Obenzoyl-2-hexadecanamido-4-octadecene-1,3-diol (17)⁶ were prepared as reported in the literature.

Scheme 3. Reagents: (a) 19, TMSOTf, AW300 molecular sieves, CH₂Cl₂, quant; (b) Zn/AcOH, 21: quant 24: 87%; (c) 22, DEPC, Et₃N, DMF, 66%; (d) 21, DEPC, Et₃N, DMF, 71%; (e) 50% TFA, 26: 86%, 28: 73%; (f) 24, DEPC, Et₃N, DMF, 79%; (g) dansyl glycine, DEPC, Et₃N, DMF, 85%; (h) NaOCH₃, 1,4-dioxane/CH₃OH, 91%.

Scheme 4. Reagents: (a) 24, DEPC, Et₃N, DMF, 83%; (b) 50% TFA, 56%; (c) dansyl glycine, DEPC, Et₃N, DMF, 94%; (d) NaOCH₃, 1,4-dioxane/CH₃OH, 91%.

3.2. 2-(Trimethylsilyl)ethyl 2-*O*-benzoyl-4,6-*O*-benzylidene-3-*O*-*p*-methoxybenzyl-1-thio-β-D-glucopyranoside (5a) and 2-(trimethylsilyl)ethyl 3-*O*-benzoyl-4,6-*O*-benzylidene-2-*O*-*p*-methoxybenzyl-β-D-glucopyranoside (5b)

A mixture of 2-(trimethylsilyl)ethyl 4,6-O-benzylideneβ-D-glucopyranoside (4) (2.9 g, 7.88 mmol), n-Bu₂SnO (2.2 g, 8.83 mmol) and 80 mL of dry benzene was stirred at reflux for 5 h. The benzene was distilled off and the stannylidene derivative was redissolved in toluene (20 mL), and *n*-Bu₄NBr (2.5 g, 7.76 mmol) and *p*-methoxybenzyl chloride (1.6 mL, 11.8 mmol) were added at 60 °C. After stirring the reaction mixture for 5 h, the solution was concentrated. Purification of the residue by column chromatography (10:1, toluene/EtOAc) on silica gel gave a mixture of 2-(trimethylsilyl)ethyl 4,6-O-benzylidene-3-O-p-methoxybenzyl-β-D-glucopyranoside and 2-(trimethylsilyl)ethyl 4,6-O-benzylidene-2-O*p*-methoxybenzyl-β-D-glucopyranoside (3.1 g, 81%); MALDIMS m/z calcd for $[C_{26}H_{36}O_7Si+Na]^+$: 511.2. Found: 511.3. To a solution of these compounds (3.1 g, 6.34 mmol) in pyridine (20 mL) was added benzoyl chloride (1.5 mL, 12.9 mmol), and the reaction mixture was stirred for 4 h at rt. Toluene was added and the solution was evaporated, the residue was extracted with CHCl₃, washed with 5% HCl, aq NaHCO₃, and H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 6:1 hexane/EtOAc as eluent to give the mixture of 5a and **5b** (3.2 g, 85%): MALDIMS m/z calcd for $[C_{33}H_{40}O_{8}]$ Si+Na]⁺: 615.3. Found: 615.5.

3.3. 2-(Trimethylsilyl)ethyl 2-*O*-benzoyl-6-*O*-benzyl-3-*O*-p-methoxybenzyl-β-D-glucopyranoside (6a)

To a solution of compounds **5a** and **5b** (2.9 g, 4.89 mmol) and sodium cyanoborohydride (2.4 g, 38.2 mmol) in dry THF (50 mL) was added powdered 3 Å molecular sieves (3 g), and the mixture was stirred

for 2 h at rt, then cooled to 0 °C. Hydrogen chloride in Et₂O was added until the solution was acidic (pH paper, gas evolution). After 1 h, the reaction mixture was poured into ice H₂O and extracted with CHCl₃. The extract was washed sequentially with aq NaHCO₃ and H₂O, dried (Na₂SO₄), and concentrated. The product was purified by silica gel column chromatography using 3:1 hexane/EtOAc as eluent to give 6a (1.24 g, 45%) and **6b** (851 mg, 29%): compound **6a**: 1 H NMR (CDCl₃): δ 8.00–6.64 (m, 14H, 3*Ph*), 5.18 (dd, 1H, $J_{1,2} = 7.9$ Hz, $J_{2.3} = 9.2 \text{ Hz}, \text{ H-2}, 4.62-4.50 (m, 4H, 2PhC}_{1}, 4.57$ (d, 1H, H-1), 3.98 (dt, 1H, CH₂CH₂Si), 3.79–3.71 (m, 3H, H-4, 6a, 6b), 3.67 (s, 3H, OCH₃), 3.61 (dd, 1H, H-3), 3.53–3.46 (m, 2H, H-5, CH₂CH₂Si), 2.81 (d, 1H, OH), 0.87-0.74 (m, 2H, CH_2CH_2Si), -0.14 (s, 9H, $Si(CH_3)_3$; ¹³C NMR (CDCl₃): δ 165.1, 159.2, 137.8, 132.9, 130.1, 129.9, 129.7, 129.6, 128.4, 127.8, 127.7, 113.7, 100.6 (C-1), 81.9 (C-3), 74.2 (C-5), 73.9 (PhCH₂), 73.7 (PhCH₂), 73.5 (C-2), 72.2 (C-4), 70.4 (C-6), 67.2 (CH_2) , 55.1 (OCH_3) , 17.9 (CH_2CH_2Si) , -1.6 $(Si(CH_3)_3)$; MALDIMS m/z calcd for $[C_{33}H_{42}O_8Si+Na]^+$: 617.3. Found: 617.4. Compound **6b**: ${}^{1}H$ NMR (CDCl₃): δ 7.96–6.58 (m, 14H, 3*Ph*), 5.18 (t, 1H, $J_{2,33,4} = 9.2$ Hz, H-3), 4.72-4.53 (m, 4H, $2PhCH_2$), 4.48 (d, 1H, $J_{1.2} = 7.3 \text{ Hz}, \text{ H-1}, 4.00 (dt, 1H, CH₂CH₂Si), 3.78–3.59$ (m, 4H, H-4, 6a, 6b, CH₂), 3.68 (s, 3H, OCH₃), 3.49 (dt, 1H, H-5), 3.44 (dd, 1H, H-2), 3.06 (d, 1H, OH), 0.71-0.60 (m, 2H, CH_2CH_2Si), -0.17 (s, 9H, $Si(CH_3)_3$); ¹³C NMR (CDCl₃): δ 167.1, 159.0, 137.8, 133.1, 130.1, 129.9, 129.8, 129.6, 128.4, 128.2, 127.8, 113.5, 103.1 (C-1), 78.0 (C-3), 77.8 (C-2), 74.3 (C-5), 73.63 (PhCH₂), 73.60 (PhCH₂), 71.0 (C-4), 70.1 (C-6), 67.6 (CH₂), 55.1 (OCH_3) , 18.5 (CH_2CH_2Si) , $-1.4 (Si(CH_3)_3)$.

3.4. Phenyl 4,6-*O*-benzylidene-3-*O*-benzyl-2-*O*-chloro-acetyl-1-thio-β-D-glucopyranoside (8)

A mixture of phenyl 4,6-*O*-benzylidene-1-thio-β-D-glucopyranoside (7) (1.0 g, 2.67 mmol), *n*-Bu₂SnO

(731 mg, 2.94 mmol), and 20 mL of dry benzene was stirred at reflux for 5 h. The benzene was distilled off and the stannylidene derivative was redissolved in toluene (10 mL) and then n-Bu₄NBr (1.29 g, 4.01 mmol) and benzyl bromide (0.48 mL, 4.04 mmol) were added at 60 °C. After the reaction mixture was stirred for 5 h, the solution was concentrated. Purification of the residue by column chromatography (20:1, toluene/EtOAc) on silica gel gave phenyl 4,6-O-benzylidene-3-O-benzyl-β-D-glucopyranoside (1.08 g, 84%); MALDIMS m/zcalcd for $[C_{26}H_{26}O_5S+Na]^+$: 473.2. Found: 473.3. To a solution of this compound (1.08 g, 2.25 mmol) in CH₂Cl₂/pyridine (4:1, 10 mL) was added chloroacetyl chloride (0.35 mL) and the mixture was stirred for 2 h at 0 °C. Toluene was added and evaporated, then extracted with CHCl₃, washed with 5% HCl, aq NaHCO₃, and H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 20:1 benzene/EtOAc as the eluent to give 8 (1.17 g, 96%): $[\alpha]_D^{24}$ +12.5 (c 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 7.49–7.24 (m, 15H, 3*Ph*), 5.57 (s, 1H, PhC*H*), 5.02 (dd, 1H, $J_{1,2} = 8.5$ Hz, $J_{2,3} = 10.3$ Hz, H-2), 4.86, 4.63 (each d, 2H, PhCH₂), 4.68 (d, 1H, H-1), 4.39 (dd, 1H, $J_{5,6a} = 10.4 \text{ Hz}$, $J_{6a,6b} = 4.7 \text{ Hz}$, H-6a), 3.94, 3.83 (each d, 2H, CH₂Cl), 3.81-3.71 (m, 3H, H-3, 4, 6b), 3.50 (dt, 1H, H-5); ¹³C NMR (CDCl₃): δ 165.7, 137.9, 137.0, 133.0, 131.5, 129.1, 129.0, 128.4, 128.3, 128.0, 127.8, 125.9, 101.2 (PhCH), 86.2 (C-1), 81.2 (C-3), 79.4 (C-4), 74.4 (PhCH₂), 72.8 (C-2), 70.5 (C-5), 68.4 (C-6), **MALDIMS** (CH₂Cl);m/zcalcd $[C_{29}H_{30}ClO_6S+Na]^+$: 564.1. Found: 564.0.

3.5. 2-(Trimethylsilyl)ethyl 4,6-*O*-benzylidene-3-*O*-benz-yl-2-*O*-chloroacetyl-β-D-glucopranosyl-(1→4)-2-*O*-benzoyl-6-*O*-benzyl-3-*O*-*p*-methoxybenzyl-β-D-glucopyranoside (9)

A solution of compound 6a (840 mg, 1.41 mmol) and 8 (1.3 g, 2.47 mmol) containing activated 4 Å molecular sieves (1.5 g) in dry CH₂Cl₂ (10 mL) was stirred under an atmosphere of argon for 2 h at rt. After cooling to -40 °C, NIS (828 mg, 3.70 mmol) and TfOH (22 μ L, 250 µmol) were successively added and stirring was continued at -40 °C for 30 min before the solution was neutralized with Et₃N. The reaction mixture was filtered, and the filtrate was washed with sodium thiosulfate and H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 20:1 toluene/EtOAc as the eluant to give 9 (810 mg, 57%). $[\alpha]_D^{24}$ +7.2 (c 0.5, CHCl₃); ¹H NMR $(CDCl_3)$: δ 7.96–6.63 (m, 24H, 5*Ph*), 5.50 (s, 1H, PhC*H*), 5.16 (t, 1H, $J_{1,2}$, $J_{2,3} = 7.9$ Hz, H-2), 4.98 (t, 1H, $J_{1',2'}$, $J_{2',3'} = 7.9 \text{ Hz}, \text{ H-2'}, 4.88-4.47 (m, 6H, 3PhC}_{2}), 4.61$ (d, 1H, $J_{1',2'} = 7.9$ Hz, H-1'), 4.47 (d, 1H, H-1), 4.23 (dd, 1H, $J_{5',6'a} = 10.6$ Hz, $J_{6'a,6'b} = 3.7$ Hz, H-6'a), 4.07 (t, 1H, H-4), 3.94 (dt, 1H, CH₂CH₂Si), 3.82, 3.77 (each,

d, 2H, CH_2Cl), 3.75–3.39 (m, 11H, H-3, H-6a, H-6b, H-6'b, 3', 4', 5', OCH_3 , CH_2CH_2Si), 3.18 (dt, 1H, H-5), 0.90–0.80 (m, 2H, CH_2CH_2Si), -0.09 (s, 9H, $Si(CH_3)_3$); ¹³C NMR (CDCl₃): δ 165.8, 165.0, 138.1, 137.9, 137.1, 132.8, 130.4, 130.2, 129.8, 129.4, 129.1, 128.5, 128.4, 128.3, 128.18, 128.15, 128.0, 127.9, 127.8, 126.0, 113.5, 101.3 (PhCH), 100.6 (C-1'), 100.1 (C-1), 81.7, 80.0, 78.2, 76.6, 75.1, 75.1, 75.0, 74.2, 74.0. 73.7, 73.3, 68.6, 67.8, 67.1, 66.0, 55.1 (OCH_3), 40.5 (CH_2Cl), 17.9 (CH_2CH_2Si), -1.5 ($Si(CH_3)_3$); MALDIMS m/z calcd for [$C_{55}H_{64}ClO_{14}Si+Na$]⁺: 1034.1. Found: 1034.0.

3.6. 2-(Trimethylsilyl)ethyl 4,6-*O*-benzylidene-3-*O*-benzyl-β-D-glucopranosyl-(1→4)-2-*O*-benzoyl-6-*O*-benzyl-3-*O*-*p*-methoxybenzoyl-β-D-glucopyranoside (10)

A solution of 9 (980 mg, 0.97 mmol), saturated NaHCO₃ in CH₃OH/H₂O (5:1 v/v) solution (48 mL), and THF (12 mL) was stirred for 1 h at 50 °C. The solution was evaporated, then extracted with CHCl3, washed with H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 60:1 toluene/EtOAc as the eluent to give 10 (700 mg, 77%): $\left[\alpha\right]_{D}^{24}$ +29.1 (c 2.0, CHCl₃); ¹H NMR (CDCl₃): δ 7.98-6.63 (m, 24H, 5Ph), 5.57 (s, 1H, PhCH), 5.22 (t, 1H, $J_{1,2}$, $J_{2,3} = 9.2 \text{ Hz}$, H-2), 4.94–4.50 (m, 6H, 3PhC H_2), 4.65 (d, 1H, $J_{1',2'} = 7.3$ Hz, H-1'), 4.51 (d, 1H, H-1), 4.16-4.11 (m, 2H, H-4, 6a), 4.00 (dd, 1H, $J_{5',6'a} = 10.6 \text{ Hz}, J_{6'a,6'b} = 3.7 \text{ Hz}, H-6'a), 3.94 (dt, 1H,$ CH_2CH_2Si), 3.82 (dd, 1H, $J_{5'.6'b} = 1.8$ Hz, H-6'b), 3.77 (dd, 1H, H-3), 3.66 (s, 3H, OCH₃), 3.61-3.49 (m, 6H, H-6b, 2', 3', 4', 5', CH_2CH_2Si), 3.19 (dt, 1H, H-5), 0.91-0.80 (m, 2H, CH₂CH₂Si), -0.09 (s, 9H, Si(CH₃)₃); ¹³C NMR (CDCl₃): δ 103.4 (C-1'), 101.1 (PhCH) 100.1 (C-1), 81.2 (C-3), 81.1 (C-3'), 80.3 (C-4'), 77.4 (C-5), 75.1 (C-2'), 74.7 (C-5 PhCH₂), 74.5 (PhCH₂), 73.5 (C-2), 68.6 (C-6), 68.2 (C-6'), 67.0 (CH₂), 66.2 (C-5), 55.0 (OCH_3) , 17.7 (CH_2CH_2Si) , $-1.4 (Si(CH_3)_3)$; MALDIMS m/z calcd for $[C_{53}H_{62}O_{13}Si+Na]^{+}$: 957.4. Found: 957.2.

3.7. 2-(Trimethylsilyl)ethyl 4,6-O-benzylidene-2-O-benzyl-3-O-benzyl- β -D-mannopyranosyl- $(1 \rightarrow 4)$ -2-O-benzyl-6-O-benzyl-3-O-p-methoxybenzyl- β -D-glucopyranoside (11)

To a solution of **10** (563 mg, 0.60 mmol) in CH_2Cl_2 (6 mL) and pyridine (3 mL), cooled to -20 °C was added triflic anhydride (610 μ L, 3.60 mmol), and the reaction mixture was stirred for 1 h at -20 °C and then for 1 h at 0 °C. Toluene was added and the solution was evaporated, then extracted with $CHCl_3$, washed with 5% HCl, aq NaHCO₃, and H₂O, dried (MgSO₄), and concentrated. The resulting residue was dissolved in DMF (6 mL) and n-Bu₄NOBz (1.75 g, 4.81 mmol), was added and the mixture was stirred for 1.5 h at rt, then for 1 h at 100 °C. Toluene was added and the solution was con-

centrated. The residue was purified by silica gel column chromatography using 10:1 toluene/EtOAc as the eluent to give 11 (501 mg 80%): $[\alpha]_D^{25}$ -15.3 (c 1.0 CHCl₃); ¹H NMR (CDCl₃): δ 8.11–6.44 (m, 29H, 6Ph), 5.70 (d, 1H, $J_{2',3'} = 3.7 \text{ Hz}$, H-2'), 5.60 (s, 1H, PhCH), 5.12 (dd, 1H, $J_{1,2} = 7.9$ Hz, $J_{2,3} = 9.2$ Hz, H-2), 4.79 (s, 1H, H-1'), 4.75–4.38 (m, 6H, 3PhCH₂), 4.42 (d, 1H, H-1), 4.30 (dd, 1H, $J_{5'.6'a} = 10.4$ Hz, $J_{6'a.6'b} = 4.3$ Hz, H-6'a), 4.14 (t, 1H, $J_{3,44,5} = 9.2 \text{ Hz}$, H-4), 4.02 (t, 1H, $J_{3',4'4',5'} = 9.2$ Hz, H-4'), 3.92 (dt, 1H, OC H_2 CH₂Si), 3.82-3.75 (m, 3H, H-6a, H-6b, H-6b), 3.63 (s, 3H, CH₃O), 3.56 (t, 1H, H-3), 3.55 (t, 1H, H-3'), 3.49–3.42 (m, 2H, H-5, OCH₂CH₂Si), 3.20 (dt, 1H, H-5'), 0.87– 0.75 (m, 2H, OCH₂CH₂Si), -0.12 (s, 9H, Si(CH₃)₃); ¹³C NMR (CDCl₃): δ 165.8, 164.9, 158.7, 137.9, 137.8, 137.4, 133.3, 132.7, 130.5, 130.2, 129.9, 129.8, 129.7, 129.4, 129.0, 128.9, 128.5, 128.4, 128.2, 127.6, 127.4, 126.1, 125.4, 113.2, 101.6 (Ph*C*H), 100.6 (C-1, $J_{C1 H1}$ = 159.3Hz), 99.4 (C-1', $J_{\text{C1.H1}} = 157.3 \text{ Hz}$), 79.9 (C-3), 78.3 (C-4), 78.0 (C-4'), 75.8 (C-3'), 75.2 (Ph*C*H₂), 74.5 (C-5), 74.0 (PhCH₂), 73.6 (PhCH₂), 73.1 (C-2), 71.5 (PhCH₂), 69.7 (C-2'), 68.7 (C-6), 68.5 (C-6'), 67.03 (C-5'), 66.98 (OCH_2CH_2Si) , 55.0, 17.8, -1.5; MALDIMS m/z calcd for $[C_{60}H_{66}O_{14}Si+Na]^+$: 1061.4. Found: 1061.2.

3.8. 2-(Trimethylsilyl)ethyl 4,6-O-benzylidene-2-O-benzyl-3-O-benzyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -2-O-benzyl- β -D-glucopyranoside (12)

To a solution of 11 (177 mg, 0.45 mmol) in CH₂Cl₂ (19 mL) and H₂O (1 mL) was added DDQ (153 mg, 0.67 mmol), and the mixture was stirred for 10 h at rt. The reaction mixture was filtered, and the filtrate was washed with H₂O, dried (MgSO₄), and concentrated. The residue was purified by silica gel column chromatography using 8:1 toluene/EtOAc as the eluent to give **12** (323 mg, 78%): $[\alpha]_{D}^{23}$ -55.7 (c 1.6, CHCl₃); ¹H NMR (CDCl₃): δ 8.07–7.15 (m, 25H, 5*Ph*,), 5.60 (s, 1H, PhCH), 5.58 (d, 1H, $J_{2',3'} = 3.7$ Hz, H-2'), 5.14 (dd, 1H, $J_{1,2} = 8.6 \text{ Hz}$, $J_{2,3} = 9.8 \text{ Hz}$, H-2), 4.79–4.47 (m, 4H, 2PhCH₂), 4.54 (s, 1H, H-1'), 4.49 (d, 1H, H-1), 4.35 (dd, 1H, $J_{5',6'a} = 10.3$ Hz, $J_{6'a,6'b} = 4.6$ Hz, H-6'a), 4.03 (t, 1H, $J_{3',4'4',5'} = 9.8$ Hz, H-4'), 3.96 (m, 1H, OCH₂CH₂Si), 3.89–3.72 (m, 6H, H-3, 4, H-6a, H-6b, H-6'b, OH), 3.60 (dd, 1H, H-3'), 3.49 (dt, 1H, OCH₂CH₂Si), 3.43 (dt, 1H, H-5), 3.37 (dt, 1H, H-5'), 0.91-0.78 (m, 2H, OCH₂CH₂Si), -0.10 (s, 9H, $Si(CH_3)_3$; ¹³C NMR (CDCl₃): δ 165.8, 165.3, 138.1, 137.7, 137.1, 133.2, 132.9, 130.1, 129.91, 129.86, 129.0, 128.5, 128.44, 128.36, 128.2, 128.1, 128.03, 127.95, 127.7, 127.5, 126.1, 101.6 (Ph*C*H), 100.5 (C-1), 100.3 (C-1'), 80.8 (C-4), 78.0 (C-4'), 75.5 (C-3'), 73.7 (C-5), 73.50 (2PhCH₂), 73.45 (C-2), 71.8 (PhCH₂), 69.2 (C-2'), 68.2 (C-3), 68.1 (C-5'), 67.1 (C-6, C-6'), 67.0 (OCH_2CH_2Si) , 17.9, -1.5; MALDIMS m/z calcd for $[C_{52}H_{58}O_{13}Si+Na]^{+}$: 941.4. Found: 941.0.

3.9. 2-(Trimethylsilyl)ethyl 4,6-O-benzylidene-2-O-benzyl-3-O-benzyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -[2,3,4-tri-O-benzyl- α -L-fucopyranosyl- $(1\rightarrow 3)$]-2-O-benzoyl-6-O-benzyl- β -D-glucopyranoside (14)

To a solution of **12** (221 mg, 0.24 mmol) and **13** (381 mg, 0.72 mmol) in dry CH₂Cl₂ (3 mL) was added powdered 4 Å molecular sieves (1 g), and the mixture was stirred for 2 h at rt, then cooled to -60 °C. NIS (241 mg, 1.08 mmol) and TfOH (6.0 μL, 72 μmol) were added to the mixture, which was stirred for 10 min at -60 °C, then neutralized with Et₃N. The solids were filtered and washed with CHCl3. The combined filtrate and washings were successively washed with aq Na₂S₂O₃ and H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 3:1 hexane/EtOAc as eluent to give 14 (317 mg, 99%): $[\alpha]_D^{24}$ -67.2 (c 1.6, CHCl₃); ¹H NMR (CDCl₃): δ 8.06-7.09 (m, 40H, 8*Ph*), 5.70 (d, 1H, $J_{2',3'} = 3.1$ Hz, H-2'), 5.52 (s, 1H, PhCH), 5.40 (t, 1H, $J_{1,22,3} = 7.9 \text{ Hz}, \text{ H-2}, 5.37 \text{ (d, } J_{1'',2''} = 3.7 \text{ Hz}, \text{ H-1''},$ 4.89 (s, 1H, H-1'), 4.88-4.17 (m, 14H, H-3, 4, H-6'a, H-5'', $5PhCH_2$), 4.40 (d, 1H, H-1), 3.97–3.78 (m, 5H, H-6a, H-6a', H-2", H-3", H-4", OCH₂CH₂Si), 3.62 (t, 1H, H-4'), 3.51-3.43 (m, 4H, H-5, H-6'b, H-3', OCH_2CH_2Si), 3.12 (dt, 1H, H-5'), 0.80–0.73 (m, 2H, $OCH_2CH_2Si)$, 0.72 (d, 3H, H-6"), -0.10 (s, 9H, $Si(CH_3)_3$); ¹³C NMR (CDCl₃): δ 166.0, 164.8, 139.2, 138.7, 138.1, 137.8, 137.7, 137.2, 133.3, 133.03, 129.94, 129.8, 129.0, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.7, 127.4, 127.2, 126.8, 126.1, 125.3, 101.4, 100.54 (C-1'), 100.46 (C-1), 95.5 (C-1"), 78.8, 78.6, 78.4, 78.3, 76.3, 75.7, 75.2, 74.9, 74.8, 73.7, 73.5, 73.0, 72.3, 71.6, 70.2, 68.9, 68.6, 67.1, 67.0, 66.1, 17.8, 16.5, -1.6; MALDIMS m/z calcd for $[C_{79}H_{86}O_{17}Si+Na]^+$: 1358.4. Found: 1358.6.

3.10. 2-(Trimethylsilyl)ethyl 3,4,6-tri-O-acetyl-2-O-benzoyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -[2,3,4-tri-O-acetyl- α -L-fucopyranosyl- $(1\rightarrow 3)$]-2-O-benzoyl-6-O-acetyl- β -D-glucopyranoside (15)

A solution of **14** (386 mg, 0.29 mmol) in THF (3.0 mL), CH₃OH (3.0 mL), and AcOH (1.0 mL) was hydrogenolysed in the presence of 10% Pd/C (400 mg) for 3 h at rt, then filtered and concentrated. The residue was acetylated with acetic anhydride (4 mL) in pyridine (6 mL). The reaction mixture was poured into ice H₂O and extracted with CHCl₃. The extract was washed sequentially with 5% HCl, aq NaHCO₃, and H₂O, dried (MgSO₄), and concentrated. The residue was purified by silica gel column chromatography using 6:1 toluene/acetone as eluent to give **15** (315 mg, quant): $[\alpha]_D^{24}$ –91.9 (*c* 1.5, CHCl₃); ¹H NMR (CDCl₃): δ 5.28 (d, $J_{1'',2''}$ = 3.1 Hz, H-1"), 4.90 (s, 1H, H-1'), 4.45 (d, 1H, $J_{1,2}$ = 7.9 Hz, H-1); ¹³C NMR (CDCl₃): δ 100.7 (C-1'),

100.1 (C-1), 95.2 (C-1"); MALDIMS m/z calcd for $[C_{51}H_{66}O_{24}Si+Na]^+$: 1114.1. Found: 1114.4.

3.11. 3,4,6-Tri-O-acetyl-2-O-benzoyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -[2,3,4-tri-O-acetyl- α -L-fucopyranosyl- $(1\rightarrow 3)$]-2-O-benzoyl-6-O-acetyl- α -D-glucopyranosyl trichloroacetimidate (16)

To a solution of **15** (320 mg, 0.29 mmol) in CH₂Cl₂ (2.0 mL), cooled to 0 °C was added CF₃CO₂H (4 mL), and the mixture was stirred for 2.5 h at rt and concentrated. EtOAc and toluene (1:2) were added and evaporated to give the reducing sugar. To a solution of the residue in CH₂Cl₂ (1.0 mL) cooled at 0 °C were added Cl₃CCN (290 μL, 2.9 mmol) and DBU (43 μL, 0.29 mmol). The reaction mixture was stirred for 2 h at 0 °C. After completion of the reaction, the mixture was concentrated. Column chromatography of the residue on silica gel (1:2, hexane/EtOAc) gave **16** (285 mg, 86%): [α]_D²⁵ –39.6 (c 1.6, CHCl₃); ¹H NMR (CDCl₃): δ 6.50 (d, 1H, J_{1,2} = 3.7 Hz, H-1), 5.43 (d, J_{1",2"} = 3.1 Hz, H-1"), 4.92 (s, 1H, H-1'); ¹³C NMR (CDCl₃): δ 100.6 (C-1'), 95.9 (C-1"), 92.9 (C-1).

3.12. 3,4,6-Tri-O-acetyl-2-O-benzoyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -[2,3,4-tri-O-acetyl- α -L-fucopyranosyl- $(1\rightarrow 3)$]-2-O-benzoyl-6-O-acetyl- β -D-glucopyranosyl- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-hexadecanamido-4-octadecene-1,3-diol (18)

To a solution of **16** (63 mg, 55 μ mol) and (2S,3R,4E)-3-O-benzovl-2-hexadecanamido-4-octadecene-1.3-diol 17 (53 mg, 83 µmol) in dry CH₂Cl₂ (1.0 mL) was added 4 Å molecular sieves (350 mg), and the mixture was stirred for 2 h at rt, then cooled to 0 °C. TMSOTf (8 µL, 44 µmol) was added, and the mixture was stirred for 24 h at 0 °C, and then neutralized with Et₃N. The solids were filtrated and washed with CHCl3. The combined filtrate and washings were successively washed with H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 3:1 toluene/EtOAc as eluent to give 18 (31 mg, 35%): $[\alpha]_{\rm D}^{25}$ -64.4 (c 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 5.28 (d, $J_{1'',2''} = 3.1 \text{ Hz}$, H-1"), 4.83 (s, 1H, H-1'), 4.40 (d, 1H, $J_{1,2} = 7.9 \text{ Hz}$, H-1); ¹³C NMR (CDCl₃): δ 100.7 (C-1'), 100.4 (C-1), 95.3 (C-1"); MALDIMS m/z calcd for $[C_{87}H_{123}NO_{27}+Na]^+$: 1636.8. Found: 1636.5.

3.13. β -D-Mannopyranosyl- $(1\rightarrow 4)$ - $[\alpha$ -L-fucopyranosyl- $(1\rightarrow 3)]$ - β -D-glucopyranosyl- $(1\rightarrow 1)$ -(2S,3R,4E)-2-hexadecanamido-4-octadecene-1,3-diol (1)

To a solution of 18 (31 mg, 19.2 μ mol) in CH₃OH (1.0 mL) and 1,4-dioxane (1.0 mL) was added NaOCH₃ (20 mg) at rt and the mixture was stirred for 24 h, then neutralized with Amberlite IR 120[H⁺]. The mixture

was filtered and concentrated. The product was purified by Sephadex LH-20 column chromatography (1:1, CHCl₃/CH₃OH) to give **1** (18 mg, 94%): $[\alpha]_D^{24}$ -8.5 (c 0.1, 1:1 CHCl₃/CH₃OH); ¹H NMR (DMSO- d_6 /D₂O, 19:1): δ 5.24 (d, 1H, $J_{1'',2''}$ = 4.3 Hz, H-1"), 4.65 (s, 1H, H-1'), 4.18 (d, 1H, $J_{1,2}$ = 7.3 Hz, H-1); MALDIMS m/z calcd for $[C_{52}H_{97}NO_{17}+Na]^+$: 1030.7. Found: 1030.2. FABMS m/z calcd for $[C_{52}H_{97}NO_{17}+Na]^+$: 1030.6654. Found: 1030.6632.

3.14. *N*-(2,2,2-Trichloroethoxycarbonyl)hexanolamine (19)

To a solution of 2,2,2-trichloroethylchloroformate (4.7 mL, 0.03 mol) in 5 mL of dioxane was added at 0 °C a mixture of hexanolamine (5 g, 0.04 mol) and MgO (3 g) in dioxane/ H_2O (1:1, 50 mL). The mixture was stirred for 16 h at rt and then EtOAc was added, the solids were filtered and washed with 5% HCl, aq NaHCO₃, and H_2O , dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography (200:1, CHCl₃/CH₃OH) to give **19** (8 g, 64%): ¹H NMR (500 MHz, CDCl₃): δ 5.03 (1H, s, N*H*), 4.72 (2H, s, C*H*₂CCl₃), 3.65 (2H, t, C*H*₂OH), 3.24 (2H, dd, NHC*H*₂), 1.61–1.36 (8H, m, 4C*H*₂); MALDIMS: m/z calcd for $[C_9H_{16}Cl_3NO_3+Na]^+$: 314.0. Found: 314.3.

3.15. 6-N-(2,2,2-Trichloroethoxycarbonyl)aminohexyl 3,4,6-tri-O-acetyl-2-O-benzoyl- β -D-mannopyranosyl-(1 \rightarrow 4)-[2,3,4-tri-O-acetyl- α -L-fucopyranosyl-(1 \rightarrow 3)-]2-O-benzoyl-6-O-acetyl- β -D-glucopyranoside (20)

To a solution of **16** (285 mg, 0.25 mmol) and **19** (95 mg, 0.33 mmol) in dry CH₂Cl₂ (2.0 mL) was added AW300 molecular sieves (500 mg), and the mixture was stirred for 2 h at rt, then cooled to 0 °C. TMSOTf (5.8 μL, 25.0 µmol) was added and the mixture was stirred for 4 h at 0 °C, then neutralized with Et₃N. The solids were filtrated and washed with CHCl₃. The combined filtrate and washings were successively washed with H₂O, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography using 5:1 toluene/acetone as eluent to give **20** (316 mg, quant): $[\alpha]_D^{25}$ -86.7 (c 1.4, CHCl₃); ¹H NMR (CDCl₃): δ 8.09–7.14 (m, 10H, 2*Ph*), 5.78 (d, 1H, $J_{2',3'} = 3.1$ Hz, H-2'), 5.51 (t, 1H, $J_{3',4'}$, $J_{4',5'} = 10.4$ Hz, H-4'), 5.37 (dd, 1H, $J_{2'',3''} = 3.1 \text{ Hz}, J_{3'',4''} = 11.0 \text{ Hz}, H-3''), 5.30 \text{ (d, 1H,}$ $J_{1'',2''} = 4.3 \text{ Hz}, \text{ H-1''}, 5.26 \text{ (t, 1H, } J_{1,2,2,3} = 8.6 \text{ Hz}, \text{ H-}$ 2), 5.23 (d, 1H, H-4"), 5.15 (dd, 1H, H-3'), 4.94 (dd, 1H, H-4"), 4.90 (s, 1H, H-1'), 4.86 (dd, 1H, H-5"), 4.78 (dd, 1H, $J_{5',6'a} = 3.1$ Hz, $J_{6'a,6'b} = 13.5$ Hz, H-6'a), 4.62 (s, 2H, CH_2CCl_3), 4.59 (dd, 1H, $J_{5,6a} = 2.5$ Hz, $J_{6a.6b} = 12.2 \text{ Hz}, \text{ H-6a}, 4.41 \text{ (dd, 1H, H-6'b)}, 4.40 \text{ (d,}$ 1H, H-1), 4.28 (dd, 1H, H-6b), 4.11 (t, 1H, $J_{3,44,5} = 9.1 \text{ Hz}, \text{ H-4}, 4.01 (t, 1H, H-3), 3.80 (dt, 1H, H-3)$

OC H_2 CH₂), 3.73 (dt, 1H, H-5'), 3.60 (dt, 1H, H-5), 3.33 (dt, 1H, OCH₂C H_2), 3.00 (dd, 2H, C H_2 NH), 2.22, 2.18, 2.04, 2.03, 2.02, 1.92, 1.91 (each s, 21H, 7Ac), 1.45–1.06 (m, 8H, 4C H_2), 0.61 (d, 3H, H-6"); ¹³C NMR (CDCl₃): δ 170.4, 170.2, 170.1, 169.8, 169.7, 169.4, 165.8, 154.4, 137.8, 133.5, 129.9, 129.6, 129.4, 129.3 129.0, 128.5, 128.4, 128.2, 125.2, 100.8 (C-1), 100.7 (C-1'), 95.7 (CCl₃), 95.2 (C-1"), 79.2 (C-3), 74.6 (C-2), 74.3 (CH₂CCl₃), 73.9 (C-4), 73.2 (C-5'), 73.1 (C-5), 71.5 (C-4"), 71.2 (C-3'), 69.9 (C-2'), 69.8 (CH₂), 67.8 (C-3"), 67.4 (C-2"), 64.7 (C-4'), 64.5 (C-5"), 62.1 (C-6'), 60.8 (C-6), 40.9, 29.2, 29.0, 26.0, 25.3, 21.4, 21.0, 20.6, 20.5, 20.4, 15.7; MALDIMS m/z calcd for [C₅₅H₆₈CINO₂₆+ Na]⁺: 1286.3. Found: 1286.7.

3.16. 6-Aminohexyl 3,4,6-tri-O-acetyl-2-O-benzoyl- β -D-mannopyranosyl- $(1\rightarrow 4)$ -[2,3,4-tri-O-acetyl- α -L-fucopyranosyl- $(1\rightarrow 3)$]-2-O-benzoyl-6-O-acetyl- β -D-glucopyranoside (21)

To a solution of 20 (317 mg, 0.25 mmol) in HOAc (5 mL) was added zinc powder (480 mg). The reaction mixture was stirred for 16 h at rt. After completion of the reaction, the mixture was filtered and the solids washed with CHCl₃. The filtrate was concentrated and purified by silica gel column chromatography (10:1, CHCl₃/CH₃OH) to give **21** (274 mg, quant): $[\alpha]_D^{23}$ –92.5 (c 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 8.08–7.16 (m, 10H, 2*Ph*), 5.77 (d, 1H, $J_{2',3'} = 3.1 \text{ Hz}$, H-2'), 5.51 (t, 1H, $J_{3',4'}$, $J_{4',5'} = 10.4 \text{ Hz}, \text{ H-4'}, 5.37 \text{ (dd, 1H, } J_{2'',3''} = 3.1 \text{ Hz},$ $J_{3'',4''} = 11.0 \text{ Hz}, \text{ H-3''}, 5.29 \text{ (d, 1H, } J_{1'',2''} = 4.3 \text{ Hz},$ H-1"), 5.26 (t, 1H, $J_{1.22.3} = 8.6$ Hz, H-2), 5.23 (d, 1H, H-4"), 5.15 (dd, 1H, H-3'), 4.93 (dd, 1H, H-4"), 4.91 (s, 1H, H-1'), 4.85 (dd, 1H, H-5"), 4.78 (dd, 1H, $J_{5'.6'a} = 3.1 \text{ Hz}, J_{6'a,6'b} = 13.5 \text{ Hz}, \text{ H-6'a}), 4.59 \text{ (dd, 1H,}$ $J_{5.6a} = 2.5 \text{ Hz}, J_{6a.6b} = 12.2 \text{ Hz}, \text{ H-6a}, 4.41 (dd, 1H, H-$ 6'b), 4.40 (d, 1H, H-1), 4.28 (dd, 1H, H-6b), 4.11 (t, 1H, $J_{3,44.5} = 9.1 \text{ Hz}, \text{ H-4}, 4.01 (t, 1H, H-3), 3.80 (dt, 1H, H-3)$ OCH_2CH_2), 3.73 (dt, 1H, H-5'), 3.61 (dt, 1H, H-5), 3.33 (dt, 1H, OCH₂CH₂), 2.62 (dd, 2H, CH₂NH), 2.23, 2.18, 2.043, 2.035, 2.02, 1.92, 1.91 (each s, 21H, 7Ac), 1.45-1.06 (m, 8H, 4C H_2), 0.61 (d, 3H, H-6"); ¹³C NMR (CDCl₃): δ 170.5, 170.4, 170.2, 169.9, 169.8, 169.5, 165.9, 164.9, 133.6, 133.5, 129.9, 129.7, 129.41, 129.36 129.0, 128.6, 128.2, 100.9 (C-1), 100.8 (C-1'), 95.3 (C-1"), 79.3, 74.7, 74.0, 73.2, 71.6, 71.3, 70.0, 69.8, 67.53, 67.47, 64.8, 64.6, 62.2, 60.9, 39.8, 29.0, 28.0, 25.9, 25.1, 21.1, 20.71, 20.65, 20.61, 20.5, 15.8; MALDIMS m/z calcd for $[C_{52}H_{68}NO_{24}+H]^+$: 1090.4. Found: 1090.8.

3.17. Syntheses of multivalent glycoconjugate building blocks

3.17.1. Compound 23. To a solution of **21** (213 mg, 0.20 mmol) and 3-(*tert*-butoxycarbonyl-carboxymethylamino)-propionic acid 2,2,2-trichloroethylester **22**

(81 mg, 0.22 mmol) in DMF (3 mL) were added Et₃N $(60 \,\mu\text{L}, 0.43 \,\text{mmol})$ and DEPC $(36 \,\mu\text{L}, 0.24 \,\text{mmol})$. The mixture was stirred for 16 h at rt. After completion of the reaction, the mixture was extracted with CHCl₃, washed with H₂O, dried (Na₂SO₄), and concentrated. The product was purified by silica gel column chromatography (3:1, toluene/acetone) to give 23 (186 mg, 66%): $\left[\alpha\right]_{D}^{23}$ -72.9 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09–7.14 (m, 10H, 2*Ph*), 6.11 (t 1H, N*H*), 5.77 (d, 1H, $J_{2',3'} = 3.1$ Hz, H-2'), 5.51 (t, 1H, $J_{3',4'}$, $J_{4',5'} = 9.8 \text{ Hz}, \text{ H-4'}, 5.37 \text{ (dd, 1H, } J_{2'',3''} = 3.0 \text{ Hz},$ $J_{3'',4''} = 11.0 \text{ Hz}, \text{ H-3''}, 5.29 \text{ (d, 1H, } J_{1'',2''} = 4.3 \text{ Hz},$ H-1"), 5.26 (t, 1H, $J_{1.22.3} = 7.9$ Hz, H-2), 5.23 (d, 1H, H-4"), 5.15 (dd, 1H, H-3'), 4.94 (dd, 1H, H-4"), 4.90 (s, 1H, H-1'), 4.87 (dd, 1H, H-5"), 4.77 (dd, 1H, $J_{5'.6'a} = 3.0 \text{ Hz}, \quad J_{6'a,6'b} = 12.8 \text{ Hz}, \quad \text{H-6'a}, \quad 4.75 \quad \text{(s, }$ 2H, CH_2CCl_3 , 4.60 (dd, 1H, $J_{5.6a} = 2.4 \text{ Hz}$, $J_{6a.6b} = 12.2 \text{ Hz}, \text{ H-6a}, 4.40 \text{ (dd, 1H, H-6b)}, 4.28 \text{ (d,}$ 1H, H-1), 4.28 (dd, 1H, H-6b), 4.11 (t, 1H, $J_{3.44.5} = 9.1 \text{ Hz}, \text{ H-4}, 4.00 (t, 1H, H-3), 3.83 (s, 2H, H-4)$ NC H_2 CO of β -alanine), 3.79 (dt, 1H, OC H_2 CH₂ of sugar unit), 3.72 (dt, 1H, H-5'), 3.61 (m, 3H, H-5, NC H_2 of β -alanine), 3.33 (dt, 1H, OC H_2 C H_2 of sugar unit), 3.00 (dd, 2H, CH_2NH of sugar unit), 2.77 (t, 2H, COCH₂ of β-alanine), 2.23, 2.18, 2.04, 2.03, 2.02, 1.92, 1.91 (each s, 21H, 7Ac), 1.47–1.06 (17H, m, t-Bu, $4(CH_2)$), 0.62 (d, 3H, H-6"); ¹³C NMR (CDCl₃): δ 170.4, 170.2, 170.1, 169.8, 169.7, 169.4, 169.0, 165.8, 164.8, 137.8, 133.5, 133.3, 129.9, 129.6, 129.33, 129.25 128.9, 128.5, 128.4, 128.1, 125.2, 100.8 (C-1), 100.7 (C-1'), 95.2 (C-1"), 94.7, 81.1, 79.2, 74.5, 74.0, 73.9, 73.13, 73.08, 71.5, 71.1, 69.9, 69.8, 67.5, 67.4, 64.7, 64.5, 62.1, 60.8, 52.4, 44.9, 39.1, 33.1, 29.1, 29.0, 28.2, 26.3, 25.3, 21.4, 21.0, 20.6, 20.4, 15.7; MALDIMS m/z calcd for $[C_{64}H_{83}Cl_3N_2O_{29}+Na]^+$: 1471.4. Found: 1471.5.

3.17.2. Compound 24. To a solution of **23** (186 mg, 0.13 mmol) in HOAc (2 mL) was added zinc powder (240 mg). The mixture was stirred for 1 h at rt and after completion of the reaction (TLC monitoring) the mixture was filtered through Celite. The filtrate was concentrated and purified by silica gel column chromatography (10:1, CHCl₃/CH₃OH) to give **24** (146 mg, 87%): $[\alpha]_D^{25}$ $-76.0 (c 1.9, CHCl_3); {}^{1}H NMR (500 MHz, CDCl_3): \delta$ 8.08–7.17 (m, 10H, 2Ph), 6.45 (t 1H, NH), 5.77 (d, 1H, $J_{2',3'} = 3.1 \text{ Hz}$, H-2'), 5.51 (t, 1H, $J_{3',4'}$, $J_{4',5'} =$ 9.8 Hz, H-4'), 5.37 (dd, 1H, $J_{2'',3''}=3.0 \text{ Hz},$ $J_{3'',4''} = 11.0 \text{ Hz}, \text{ H-3''}, 5.30 \text{ (d, 1H, } J_{1'',2''} = 3.7 \text{ Hz},$ H-1"), 5.26 (t, 1H, $J_{1,22,3} = 7.9$ Hz, H-2), 5.23 (d, 1H, H-4"), 5.16 (dd, 1H, H-3'), 4.94 (dd, 1H, H-4"), 4.91 (s, 1H, H-1'), 4.86 (dd, 1H, H-5"), 4.77 (dd, 1H, $J_{5',6'a} = 3.0 \text{ Hz}, \quad J_{6'a,6'b} = 12.8 \text{ Hz}, \quad \text{H-6'a}), \quad 4.60 \quad (dd,$ 1H, $J_{5,6a} = 2.4$ Hz, $J_{6a,6b} = 12.2$ Hz, H-6a), 4.40 (dd, 1H, H-6'b), 4.28 (d, 1H, H-1), 4.28 (dd, 1H, H-6b), 4.11 (t, 1H, $J_{3.44.5} = 9.1$ Hz, H-4) 4.01 (t, 1H, H-3), 3.88 (s, 2H, NC H_2 CO of β -alanine), 3.79 (dt, 1H,

OC H_2 CH₂ of sugar unit), 3.72 (dt, 1H, H-5'), 3.61 (m, 3H, H-5, NC H_2 of β-alanine), 3.33 (dt, 1H, OCH₂C H_2 of sugar unit), 3.06 (dd, 2H, C H_2 NH of sugar unit), 2.57 (t, 2H, COCH₂ of β-alanine), 2.23, 2.18, 2.04, 2.03, 2.02, 1.92, 1.91 (each s, 21H, 7Ac), 1.47–1.06 (17H, m, t-Bu, 4(C H_2)), 0.62 (d, 3H, H-6"); ¹³C NMR (CDCl₃): δ 170.44, 170.36, 170.1, 169.9, 169.7, 169.4, 165.8, 165.0, 133.5, 133.4, 129.9, 129.6, 129.33, 129.2, 128.5, 128.4, 100.8 (C-1), 100.7 (C-1'), 95.2 (C-1"), 79.1, 77.2, 74.6, 73.9, 73.13, 73.08, 71.4, 71.2, 69.9, 69.8, 67.44, 67.38, 64.7, 64.5, 62.1, 60.8, 39.3, 29.0, 28.1, 26.2, 25.3, 21.0, 20.6, 20.54, 20.51, 20.41, 15.7; MALDIMS m/z calcd for [C₆₂H₈₂N₂O₂₉+Na]⁺: 1341.5. Found: 1341.9.

3.18. Syntheses of dimeric glycoconjugate

3.18.1. Compound 25. To a solution of **24** (71 mg, 54 μmol) and **21** (65 mg, 59 μmol) in DMF (1.5 mL) were added Et₃N (15 μL, 108 μmol) and DEPC (9 μL, 59 µmol). The mixture was stirred for 16 h at rt. After completion of the reaction, the mixture was extracted with EtOAc, washed with H₂O, dried (Na₂SO₄), and concentrated. The product was purified by silica gel column chromatography (20:1, CHCl₃/CH₃OH) to give 25 (92 mg, 71%): $[\alpha]_D^{23} - 98.0$ (c 1.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09–7.29 (m, 20H, 4*Ph*), 5.78 (d, 2H, $J_{2',3'} = 3.1 \text{ Hz}$, H-2'), 5.51 (t, 2H, $J_{3',4'}$, $J_{4'.5'} = 9.8 \text{ Hz}, \text{ H-4'}, 5.37 \text{ (dd, 2H, } J_{2''.3''} = 3.2 \text{ Hz},$ $J_{3'',4''} = 11.0 \text{ Hz}, \text{ H-3''}, 5.29 \text{ (d, 2H, } J_{1'',2''} = 3.7 \text{ Hz}, \text{ H-}$ 1"), 5.26 (t, 2H, $J_{1,2,2,3} = 7.9$ Hz, H-2), 5.23 (d, 2H, H-4"), 5.15 (dd, 2H, H-3'), 4.94 (dd, 2H, H-4"), 4.91 (s, 2H, H-1'), 4.87 (dd, 2H, H-5"), 4.77 (dd, 2H, $J_{5',6'a} = 3.0 \text{ Hz}, J_{6'a,6'b} = 12.8 \text{ Hz}, H-6'a), 4.60 \text{ (br dd,}$ 2H, $J_{5,6a} = 2.4 \text{ Hz}$, $J_{6a,6b} = 12.8 \text{ Hz}$, H-6a), 4.40 (d, 2H, H-1), 4.39 (dd, 2H, H-6'b), 4.28 (dd, 2H, H-6b), 4.11 (t, 2H, $J_{3,44,5} = 9.1$ Hz, H-4) 4.00 (t, 2H, H-3), 3.81–3.73 (m, 6H, 2H-5', 2NC H_2 CO of β -alanine, 2OCH₂CH₂ of sugar unit), 3.61 (br dd, 2H, H-5), 3.56 (br t, 2H NC H_2 of β -alanine), 3.33 (dt, 2H, OC H_2 C H_2 of sugar unit), 3.00 (dd, 4H, CH₂NH of sugar unit), 2.37 (t, 2H, $COCH_2$ of β -alanine), 2.22, 2.18, 2.05, 2.04, 2.02, 1.92, 1.91 (each s, 42H, 14Ac), 1.47-1.06 $(25H, m, t-Bu, 8(CH_2)), 0.62 (d, 6H, H-6'');$ ¹³C NMR $(CDCl_3)$: δ 170.4, 170.2, 170.1, 169.8, 169.7, 169.4, 169.0, 165.8, 164.9, 133.5, 133.3, 129.9, 129.6, 129.3, 129.2, 128.5, 128.4, 100.8 (C-1), 100.7 (C-1'), 95.2 (C-1"), 80.6, 79.2, 77.3, 74.5, 73.9, 73.12, 73.07, 71.4, 71.1, 69.9, 67.4, 67.3, 64.7, 64.5, 62.1, 60.8, 39.4, 39.1, 29.2, 26.3, 25.3, 21.0, 20.6, 20.54, 20.51, 15.7; MALDIMS m/z calcd for $[C_{114}H_{147}N_3O_{52}+Na]^+$: 2412.9. Found: 2413.3.

3.18.2. Compound 26. To a solution of **25** (90 mg, 37.6 μ mol) in CH₂Cl₂ (1 mL) was added CF₃CO₂H (1 mL) and the mixture was stirred for 3 h at rt. After

completion of the reaction, the mixture was concentrated and purified by silica gel column chromatography (10:1, CHCl₃/CH₃OH) to give **26** (74 mg, 86%). MALD-IMS m/z calcd for $[C_{109}H_{139}N_3O_{50}+Na]^+$: 2312.8. Found: 2313.4.

3.19. Syntheses of trimeric glycoconjugate

3.19.1. Compound 27. To a solution of **26** (70 mg. 30.6 μmol) and **24** (44 mg, 33.7 μmol) in DMF (1 mL) were added Et₃N (9.1 μL, 66.0 μmol) and DEPC (5.5 μL, 36.0 μmol). The mixture was stirred for 16 h at rt. After completion of the reaction, the mixture was extracted with CHCl₃, washed with H₂O, dried (Na₂SO₄), and concentrated. The product was purified by silica gel column chromatography (30:1, CHCl₃/ CH₃OH) to give **27** (85 mg, 79%): $[\alpha]_D^{23}$ -98.0 (c 2.6, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09–7.33 (m, 30H, 6*Ph*), 5.78 (d, 3H, $J_{2',3'} = 3.1$ Hz, H-2'), 5.51 (t, 3H, $J_{3',4'}$, $J_{4',5'} = 10.4$ Hz, H-4'), 5.37 (dd, 3H, $J_{2'',3''} = 3.7 \text{ Hz}, J_{3'',4''} = 10.9 \text{ Hz}, H-3''), 5.29 \text{ (d, } 3H,$ $J_{1'',2''} = 3.7 \text{ Hz}, \text{ H-1''}, 5.25 \text{ (t, 3H, } J_{1,2,2,3} = 7.9 \text{ Hz, H-1}$ 2), 5.23 (d, 3H, H-4"), 5.15 (dd, 3H, H-3'), 4.95 (dd, 3H, H-4"), 4.93 (s, 3H, H-1'), 4.87 (dd, 3H, H-5"), 4.77 (dd, 3H, $J_{5',6'a} = 3.0$ Hz, $J_{6'a,6'b} = 12.8$ Hz, H-6'a), 4.60 (br dd, 3H, $J_{5,6a} = 2.4$ Hz, $J_{6a,6b} = 12.8$ Hz, H-6a), 4.43 (d, 3H, H-1), 4.42 (dd, 3H, H-6'b), 4.28 (dd, 3H, H-6b), 4.12 (t, 3H, $J_{3,44,5} = 9.1$ Hz, H-4) 4.00 (t, 3H, H-3), 3.83–3.75 (m, 8H, 3H-5', 2NC H_2 CO of β-alanine, $3OCH_2CH_2$ of sugar unit), 3.67-3.57 (m, 5H, 3H-5, 2NC H_2 of β -alanine), 3.33 (dt, 3H, OCH₂C H_2 of sugar unit), 3.00 (dd, 6H, CH_2NH of sugar unit), 2.37 (t, 2H, $COCH_2$ of β -alanine), 2.23, 2.18, 2.05, 2.04, 2.02, 1.92, 1.91 (each s, 63H, 21Ac), 1.34-1.05 (33H, m, t-Bu, $12(CH_2)$), 0.62 (d, 9H, H-6"); ¹³C NMR (CDCl₃): δ 170.3, 170.2, 169.9, 169.7, 169.6, 169.3, 165.7, 164.7, 162.4, 133.4, 133.2, 129.8, 129.5, 129.20, 129.17, 128.4, 128.3, 100.8 (C-1), 100.5 (C-1'), 95.1 (C-1"), 79.1, 77.2, 74.5, 73.8, 73.0, 71.3, 71.1, 69.8, 67.34, 67.26, 64.6, 64.4, 62.0, 60.7, 39.2, 36.3, 31.2, 28.9, 28.1, 26.3, 25.2, 20.8, 20.5, 20.44, 20.39, 20.3, 15.5; MALDIMS m/z calcd for $[C_{171}H_{219}N_5O_{78}+Na]^+$: 3613.3. Found: 3613.0.

3.19.2. Compound 28. Compound **28** was derived from **27** according to the procedure described for the synthesis of **26**; yield: 60 mg (73%): MALDIMS m/z calcd for $[C_{166}H_{211}N_5O_{76}+Na]^+$: 3513.3. Found: 3512.8.

3.19.3. Compound 29. To a solution of **28** (24 mg, 6.88 μ mol) and dansyl glycine (6.3 mg, 20.6 μ mol) in DMF (1 mL) were added Et₃N (5.2 μ L, 38.0 μ mol) and DEPC (3.1 μ L, 20.0 μ mol) and the mixture was stirred for 16 h at rt. After completion of the reaction, the mixture was extracted with CHCl₃, washed with H₂O, dried (Na₂SO₄), and concentrated. The product was purified by silica gel column chromatography (30:1,

CHCl₃/CH₃OH) to give dansyl derivative 29 (22 mg, 85%): $[\alpha]_D^{23}$ -74.4 (c 0.7, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09–7.27 (m, 36H, C₁₀H₆ of dansyl glycine, 6*Ph*), 5.29 (d, 3H, $J_{1'',2''} = 3.7$ Hz, H-1"), 4.93 (s, 3H, H-1'), 4.40 (d, 3H, $J_{1,2} = 7.9$ Hz, H-1), 3.97–3.54 (m, 15H, 3H-5, 3H-5', 2NCH₂CO of β-alanine, 3OCH₂CH₂ of sugar unit, 2NCH₂ of β-alanine, NHCH₂CO of dansyl glycine), 3.33 (dt, 3H, OCH₂CH₂ of sugar unit), 3.00 (dd, 6H, CH_2NH of sugar unit), 2.89 (6H, s, $N(CH_3)_2$ of dansyl glycine), 2.77 (t, 2H, $COCH_2$ of β -alanine), 2.23, 2.18, 2.05, 2.04, 2.02, 1.92, 1.91 (each s, 63H, 21Ac), 1.34–1.05 (24H, m, 12(C H_2)), 0.61 (d, 9H, H-6"); ¹³C NMR (CDCl₃): δ 170.5, 170.3, 170.1, 169.9, 169.7, 169.4, 165.8, 164.9, 133.6, 133.4, 129.9, 129.6, 129.3, 128.9, 128.5, 100.9 (C-1), 100.7 (C-1'), 95.3 (C-1"), 79.2, 74.6, 74.0, 73.1, 71.5, 71.2, 70.0, 67.5, 67.4, 64.7, 64.5, 62.2, 60.9, 45.4, 39.4, 28.9, 26.4, 25.3, 21.0, 20.7, 20.5, 15.7; MALDIMS m/z calcd $[C_{180}H_{225}N_7O_{79}S+Na]^+$: 3802.4. Found: 3802.7.

3.19.4. Compound **2.** Compound **2** was synthesized from 29 according to the procedure described for the synthesis of 1; yield: 11 mg (91%): $[\alpha]_D^{23}$ -61.3 (c 0.4, CH₃OH/H₂O 1:1); ¹H NMR (500 MHz, 1:1 CD₃OD/ D_2O): δ 8.54–7.35 (m, 6H, $C_{10}H_6$ of dansyl glycine), 5.46 (d, 3H, $J_{1'',2''} = 3.7$ Hz, H-1"), 4.56 (br d, 3H, H-1'), 4.36 (br d, 3H, H-1), 2.88, 2.87 (6H, each s, $N(CH_3)_2$ of dansyl glycine), 1.18 (d, 9H, H-6"); ¹³C NMR (125 MHz, 1:1 CD₃OD/D₂O): δ 174.0, 171.1, 170.8, 170.1, 152.4, 135.5, 131.3, 130.5, 130.4, 129.6, 124.7, 120.2, 120.1, 116.8, 103.5, 100.3, 99.8, 79.5, 78.6, 78.1, 76.4, 75.7, 75.3, 74.1, 73.3, 72.9, 71.7, 71.3, 71.1, 70.6, 69.8, 68.3, 68.1, 62.7, 61.8, 61.5, 52.5, 49.4, 45.9, 45.4, 44.9, 40.4, 30.0, 29.6, 27.1, 26.1, 16.7; MALDIMS m/z calcd for $[C_{96}H_{159}N_7O_{52}S+Na]^+$: 2298.0. Found: 2297.6.

3.20. Syntheses of tetrameric glycoconjugate

3.20.1. Compound 30. Compound **30** was synthesized from 28 according to the procedure described for the synthesis of **25**; yield: 42 mg (83%): $[\alpha]_D^{23}$ -99.0 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.08–7.27 (m, 40H, 8*Ph*), 5.78 (d, 4H, $J_{2',3'} = 3.1$ Hz, H-2'), 5.51 (t, 4H, $J_{3',4'}$, $J_{4',5'} = 10.4$ Hz, H-4'), 5.37 (dd, 4H, $J_{2'',3''} =$ 3.7 Hz, $J_{3'',4''} = 10.9$ Hz, H-3"), 5.29 (d, 4H, $J_{1'',2''} = 3.7 \text{ Hz}, \text{ H-1''}, 5.28 \text{ (t, 4H, } J_{1,22,3} = 7.9 \text{ Hz}, \text{ H-}$ 2), 5.23 (d, 4H, H-4"), 5.15 (dd, 4H, H-3'), 4.95 (dd, 4H, H-4"), 4.91 (s, 4H, H-1'), 4.85 (dd, 4H, H-5"), 4.77 (dd, 4H, $J_{5',6'a} = 3.0$ Hz, $J_{6'a,6'b} = 12.8$ Hz, H-6'a), 4.60 (br dd, 4H, $J_{5,6a} = 2.4 \text{ Hz}$, $J_{6a,6b} = 12.8 \text{ Hz}$, H-6a), 4.41 (d, 4H, H-1), 4.40 (br d, 4H, H-6'b), 4.28 (br d, 4H, H-6b), 4.11 (t, 4H, $J_{3,44,5} = 9.1$ Hz, H-4) 4.00 (t, 4H, H-3), 3.83-3.75 (m, 10H, 4H-5', 2NCH₂CO of β -alanine, 4OC H_2 CH₂ of sugar unit), 3.67–3.57 (m, 6H, 4H-5, 2NC H_2 of β -alanine), 3.33 (dt, 4H, OCH₂CH₂ of sugar unit), 3.00 (dd, 8H, CH₂NH of sugar unit), 2.37 (t, 2H, COCH₂ of β-alanine), 2.22, 2.18, 2.04, 2.03, 2.02, 1.92, 1.91 (each s, 84H, 28Ac), 1.35–1.04 (41H, m, *t*-Bu, 16(CH₂)), 0.62 (d, 12H, H-6"); ¹³C NMR (CDCl₃): δ 170.5, 170.3, 170.1, 169.8, 169.7, 169.4, 165.8, 164.9, 133.6, 133.4, 129.9, 129.6, 129.4, 129.3, 128.6, 128.5, 100.9 (C-1), 100.7 (C-1'), 95.3 (C-1"), 79.2, 74.6, 74.0, 73.2, 73.1, 71.5, 71.2, 70.0, 67.5, 67.4, 64.7, 64.5, 62.2, 60.9, 39.4, 39.2, 28.22, 28.17, 26.5, 26.4, 25.4, 21.0, 20.7, 20.61, 20.57, 20.5, 15.7; MALDIMS m/z calcd for [C₂₂₈H₂₉₁N₇O₁₀₄+Na]⁺: 4813.8. Found: 4814.0.

3.20.2. Compound 31. Compound **31** was synthesized from **30** according to the procedure described for the synthesis of **26**; yield: 24 mg (56%); MALDIMS m/z calcd for $[C_{223}H_{283}N_7O_{102}+Na]^+$: 4713.7. Found: 4713.9.

3.20.3. Compound 32. Compound **32** was synthesized from 31 according to the procedure described for the synthesis of **29**; yield: 24 mg (94%): $[\alpha]_D^{23}$ -85.0 (c 0.8, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09–7.27 (m, 46H, $C_{10}H_6$ of dansyl glycine, 8Ph), 5.28 (d, 4H, $J_{1'',2''} = 3.7 \text{ Hz}, \text{ H-1''}, 4.92 \text{ (s, 4H, H-1')}, 4.42 \text{ (d, 4H, H-1')}$ $J_{1,2} = 7.9 \text{ Hz}, \text{ H-1}, 3.97-3.54 (m, 20H, 4H-5, 4H-5')$ 2NCH₂CO of β-alanine, 4OCH₂CH₂ of sugar unit, 4NC H_2 of β -alanine, NHC H_2 CO of dansyl glycine), 3.33 (dt, 4H, OCH₂CH₂ of sugar unit), 3.00 (dd, 8H, CH_2NH of sugar unit), 2.89 (6H, s, $N(CH_3)_2$ of dansyl glycine), 2.77 (t, 2H, COC H_2 of β -alanine), 2.23, 2.18, 2.05, 2.04, 2.02, 1.92, 1.91 (each s, 84H, 28Ac), 1.34-1.05 (32H, m, $16(CH_2)$), 0.61 (d, 12H, H-6"); ^{13}C NMR (CDCl₃): δ 170.5, 170.3, 170.1, 169.9, 169.7, 169.5, 165.8, 164.9, 133.6, 133.4, 129.9, 129.6, 129.3, 128.6, 128.5, 100.9 (C-1), 100.7 (C-1'), 95.3 (C-1"), 79.2, 74.6, 74.0, 73.1, 71.5, 71.2, 70.0, 67.5, 67.4, 64.7, 64.5, 62.2, 60.9, 39.4, 29.7, 29.2, 29.1, 26.41, 26.35, 25.4, 21.0, 20.7, 20.6, 20.5, 15.7; MALDIMS m/z calcd for $[C_{237}H_{297}N_9O_{105}S+Na]^+$: 5003.4. Found: 5003.9.

3.20.4. Compound 3. Compound **3** was synthesized from **32** according to the procedure described for the synthesis of **2**; yield: 13.6 mg (91%): $\left[\alpha\right]_D^{23}$ –55.8 (c 0.5, CH₃OH/H₂O 1:1); ¹H NMR (500 MHz, CDCl₃): δ 8.39–7.28 (m, 6H, C₁₀H₆ of dansyl glycine), 5.42 (br t, 4H, H-1"), 4.53 (br d, 4H, H-1'), 4.33 (d, 4H, $J_{1,2}=7.9$ Hz, H-1), 2.84, 2.83 (6H, each s, N(CH₃)₂ of dansyl glycine), 1.18 (d, 9H, H-6"); ¹³C NMR (125MHz, 1:1 CD₃OD/D₂O): δ 103.4, 100.4, 99.7, 79.3, 79.0, 78.0, 76.4, 75.7, 75.3, 74.4, 73.3, 72.9, 71.7, 71.3, 70.6, 69.7, 68.3, 68.1, 62.7, 61.7, 61.5, 46.0, 40.4, 30.0, 29.6, 27.2, 27.1, 26.1, 16.7; MALDIMS m/z calcd for $\left[C_{125}H_{209}N_9O_{69}S+Na\right]^+$: 2995.3. Found: 2995.0.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.carres. 2006.04.019.

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